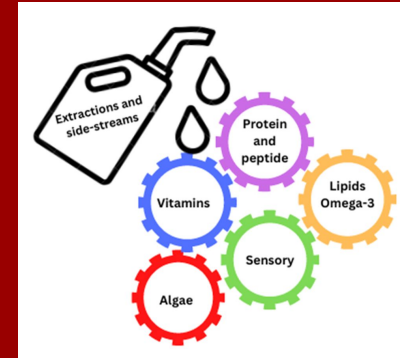


# Recent Trends in Lipid Oxidation and Antioxidants in Omega-3 PUFA Rich Food Emulsions

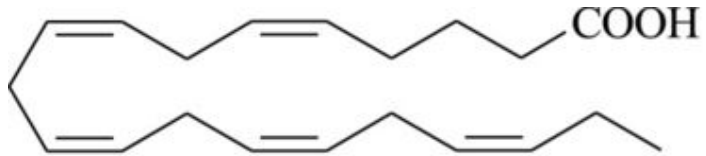
Charlotte Jacobsen  
Professor and group leader  
Bioactives – Analysis and Applications  
National Food Institute  
Technical University of Denmark  
[chja@food.dtu.dk](mailto:chja@food.dtu.dk)



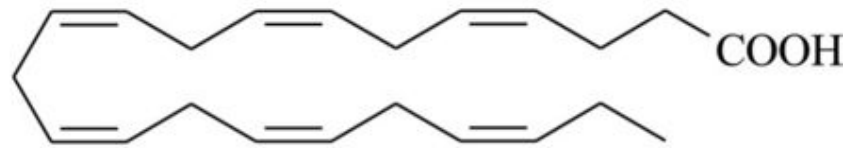
# Outline

- Introduction to omega-3 PUFA
- Introduction to lipid oxidation
- Factors affecting lipid oxidation in food emulsions
  - Role of interface and emulsifier
- Peptides and protein hydrolysates as antioxidants – a new approach
- Effect of peptides in emulsions at different pH
- Screening of metal chelating peptides
- Conclusions

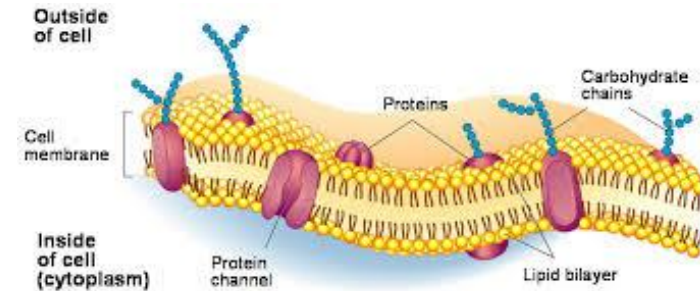
# Health effects of omega-3 PUFA



Eicosapentaenoic acid  
(EPA,  $C_{20}H_{30}O_2$ )

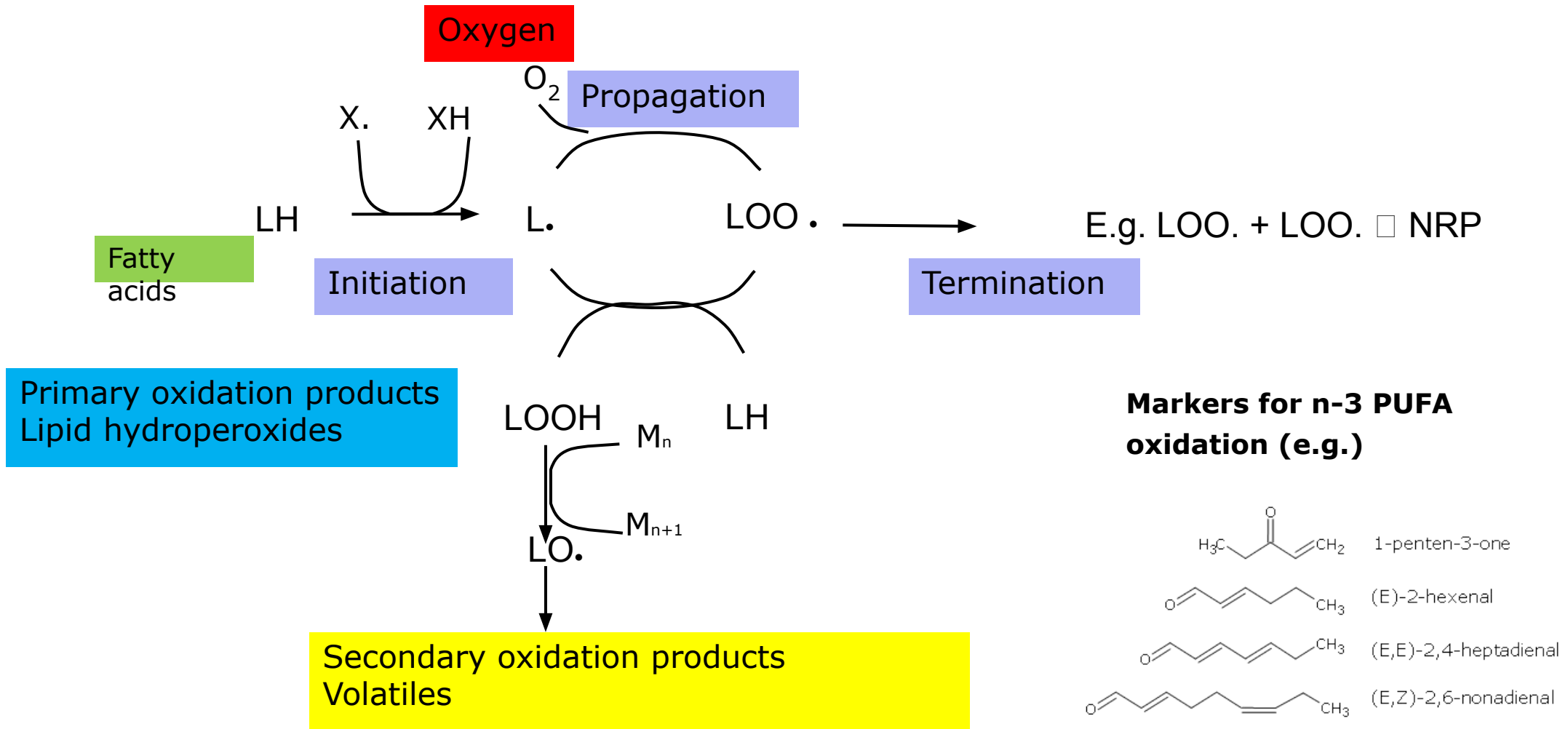


Docosahexaenoic acid  
(DHA,  $C_{22}H_{32}O_2$ )

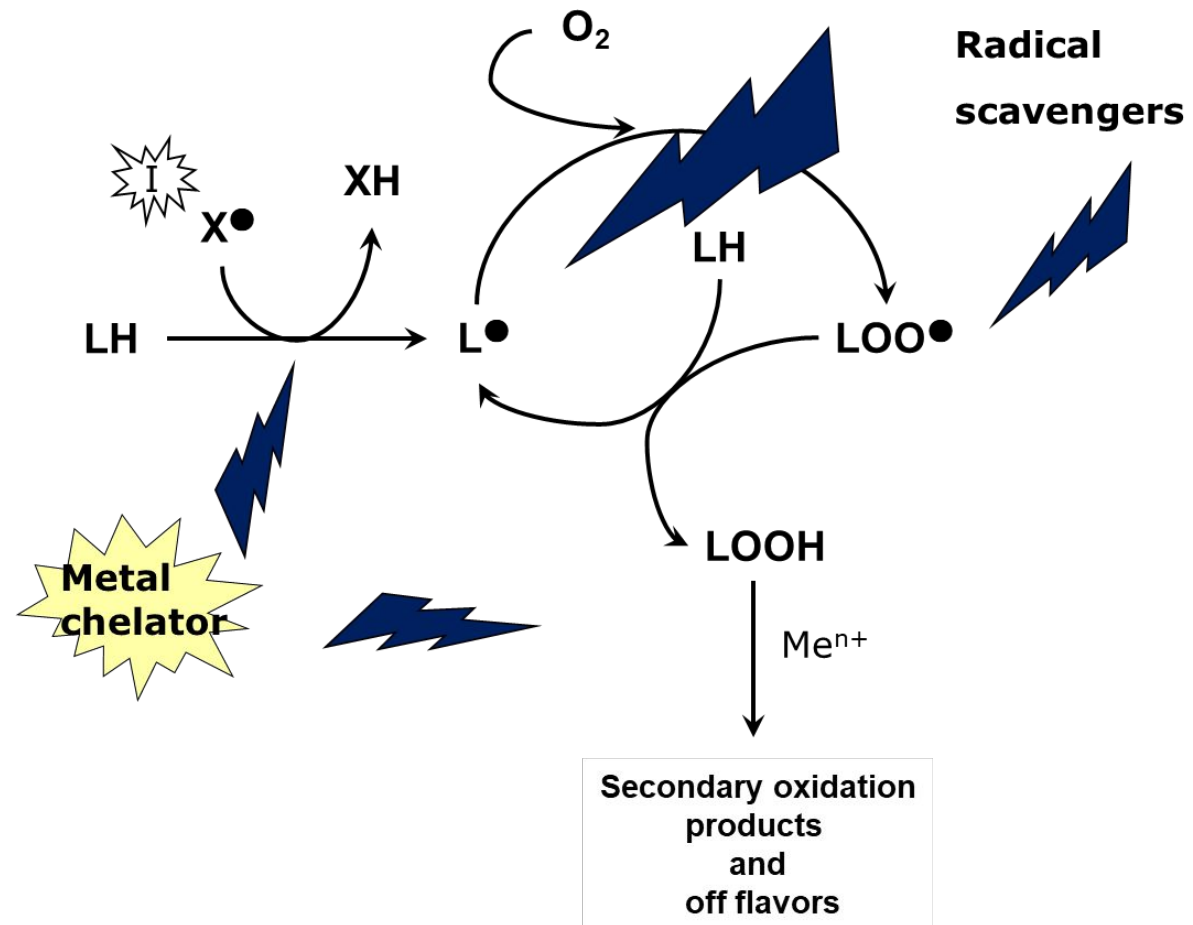


*Prof. Calder: "We may need at least 1 g/day of EPA and DHA"*  
*European Lipid Science Award lecture 2021*

# Lipid oxidation reactions: The basic reactions



# Antioxidant mechanisms



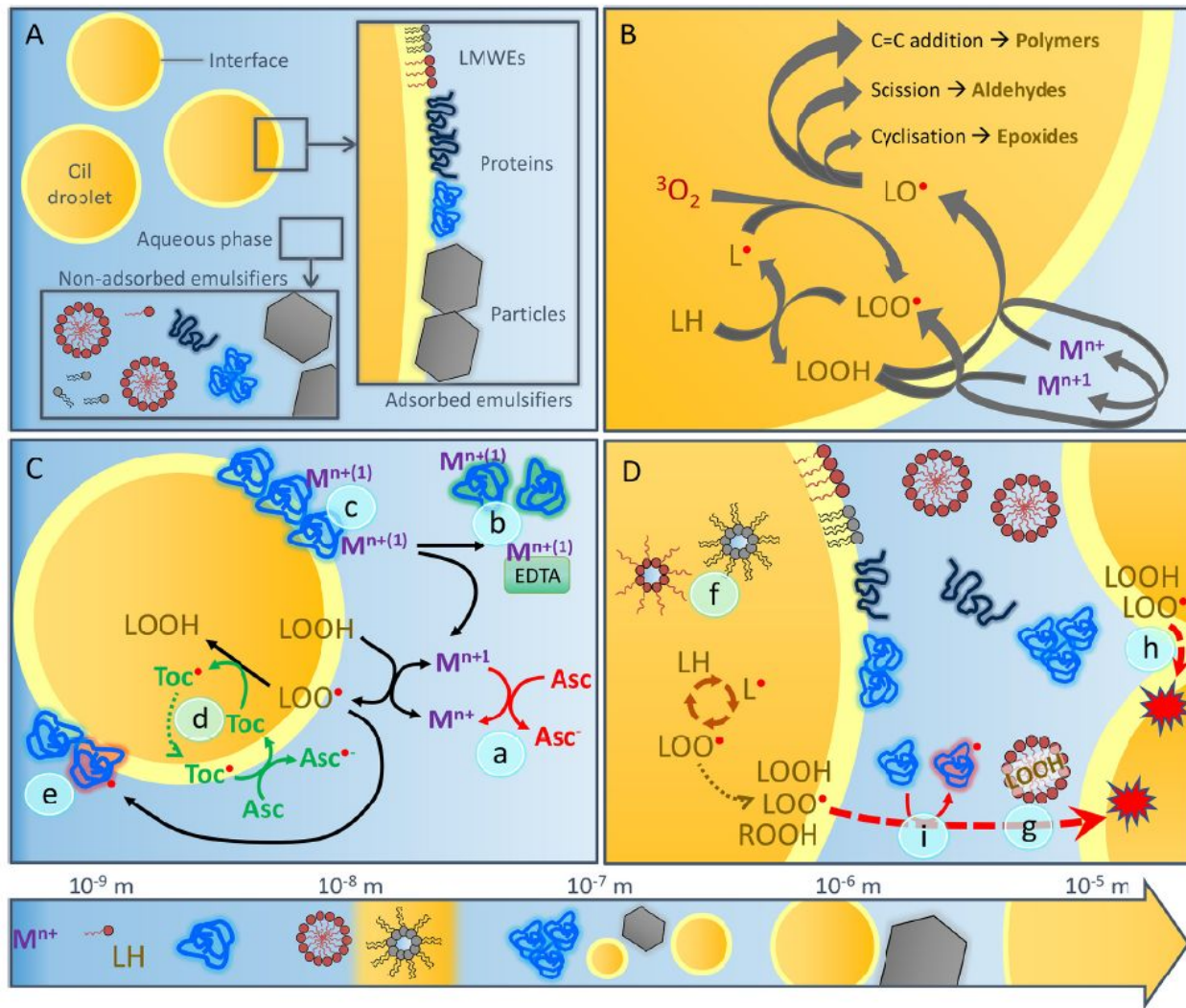


Fig. 1. (A) General structure of an O/W emulsion and of various types of food emulsifiers. (B) Simplified overview of dominant chemical pathways of lipid oxidation in emulsions, with an emphasis on the redox cycling engine involving transition metals (reduced form  $M^{n+}$ , oxidized form  $M^{n+1}$ ). (C) Pro- (red) and antioxidant (green) mechanisms as influenced by the surface of oil droplets. (D) Possible transport routes (red arrows) of lipid oxidation intermediates (e.g., lipid oxidation products bearing a hydroperoxide group) between emulsion droplets. LH, unsaturated lipid;  $L^\bullet$ , alkyl radical;  $LOO^\bullet$ , peroxy radical; LOOH, lipid hydroperoxide;  $LO^\bullet$ , alkoxy radical; ROOH, molecule bearing a hydroperoxide group (e.g., hydroperoxy-alkenal); Asc, ascorbate; Toc, tocopherol. The letters (a) to (i) point to mechanisms referred to and further explained in the text. The large arrow at the bottom depicts typical dimensions (m) of the molecules and structures discussed. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

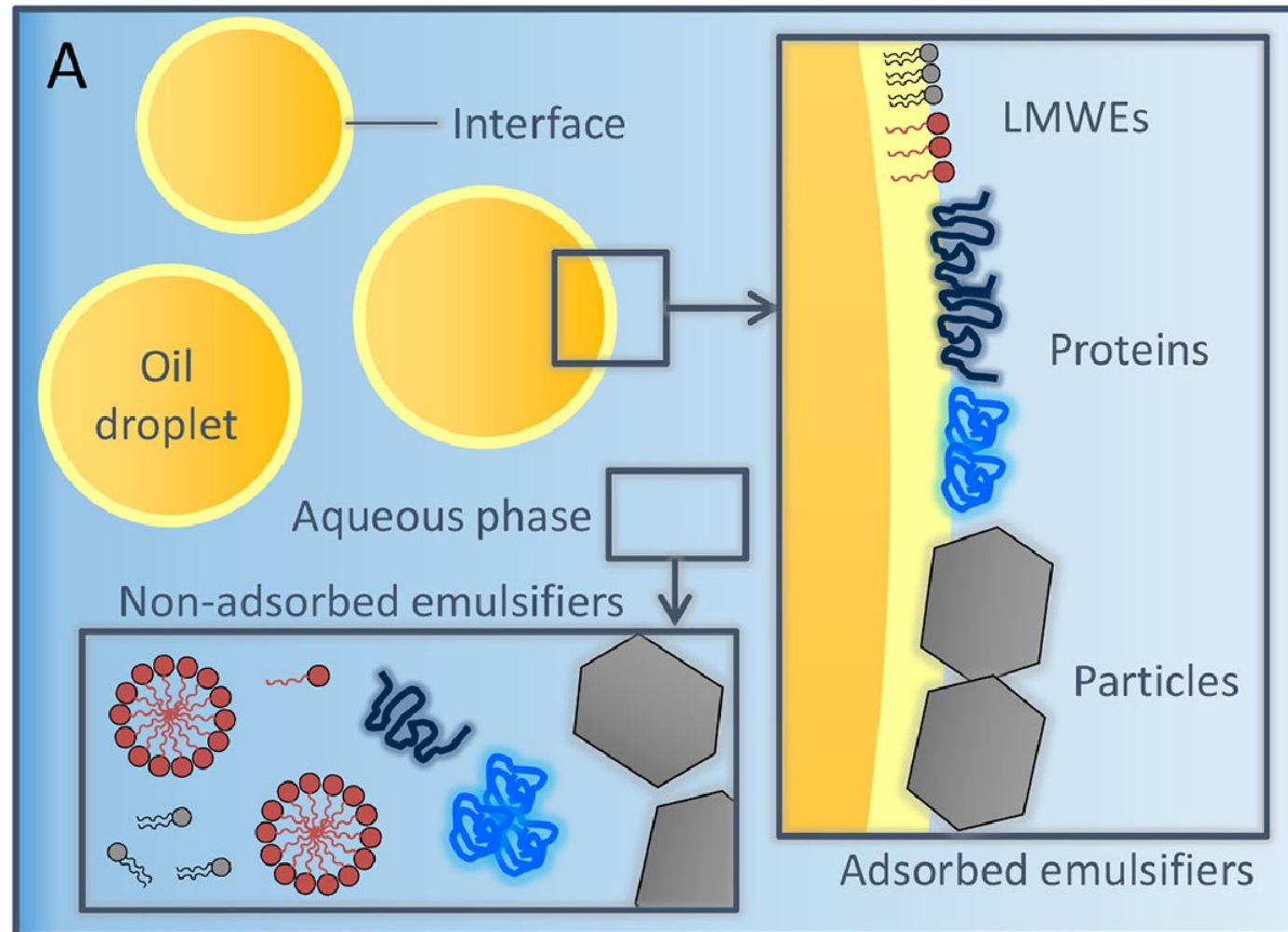
## Aspects of lipid oxidation in emulsions

Berton-Carabin, C., Villeneuve, P., Durand, E., Lecomte, J., van Duynhoven, J., Meynier, A., Yesiltas, B., Jacobsen, C., Hennebelle, M. (2024).

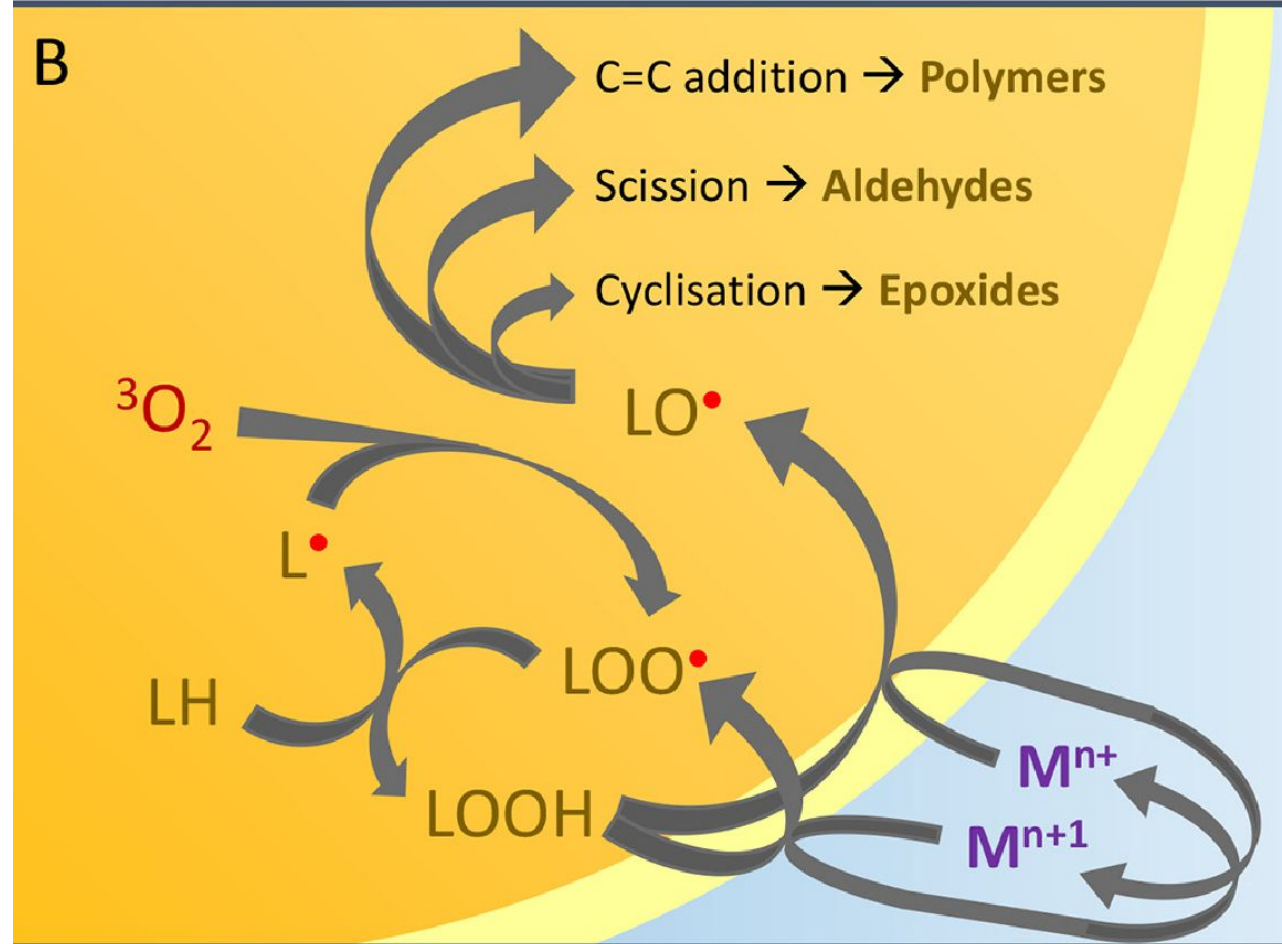
Lipid oxidation in emulsions: New insights from the past two decades. *Progress Lipid Res.* 94, 101275



# Structure of emulsions and types of emulsifiers



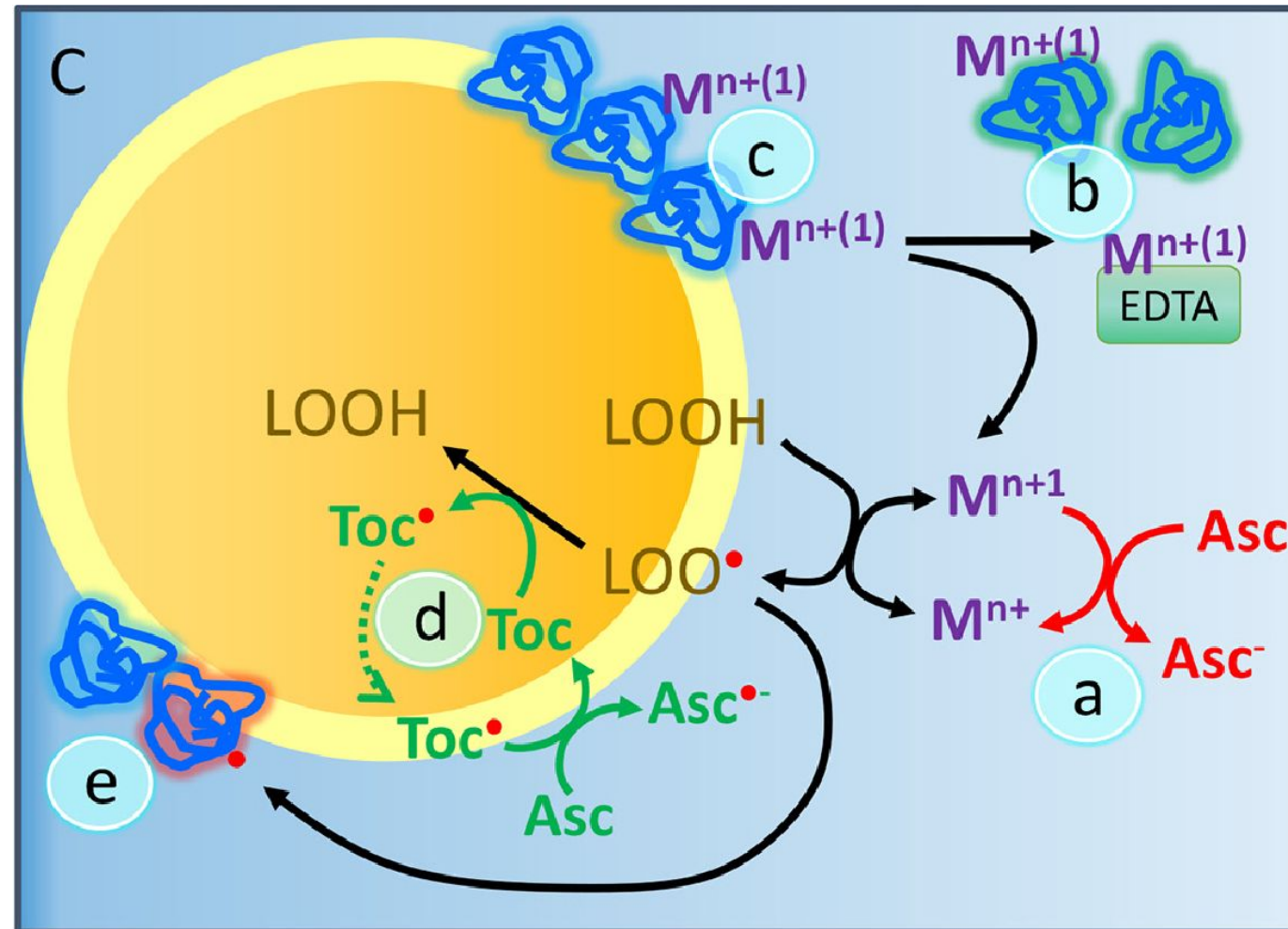
# Location of chemical pathways of lipid oxidation in emulsions





# Pro- (red) and antioxidant (green) mechanisms as influenced by the surface of oil droplets

- a: Reduction of metal ions by Asc
- b and c: Binding of metal ions at the interface or in the aqueous phase
- d: Tocopherol as an antioxidant and regeneration by Asc
- e: Co-oxidation of proteins



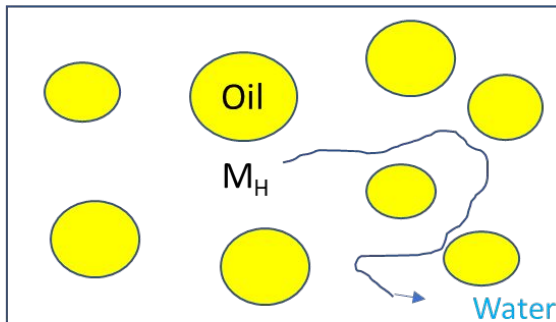
# The role of mass transport in lipid oxidation

How does lipid oxidation spread within an emulsion ? How do lipid oxidation reactants transfer from lipid droplets to lipid droplets ?

## MASS TRANSPORT PATHWAYS

### Pathway A:

Aqueous diffusion of hydrophilic molecules

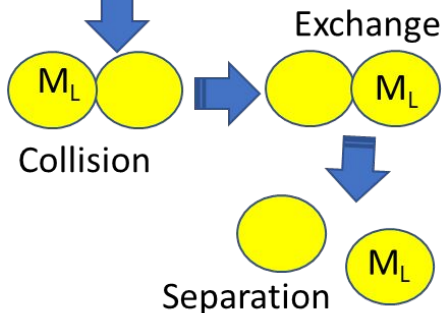
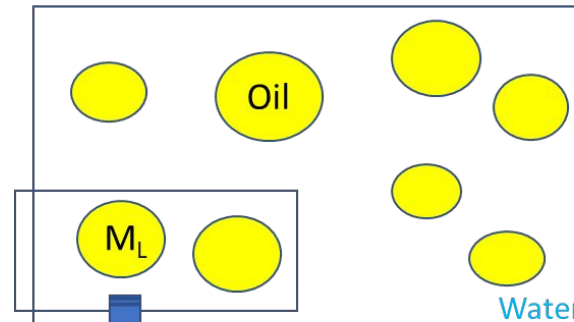


$M_H$  : Hydrophilic molecule

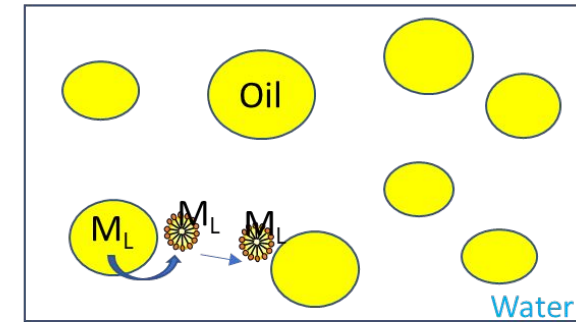
$M_L$  : Lipophilic molecule

### Pathway B:

Inter droplets transfer (collision/exchange/separation) of lipophilic molecules



### Pathway c: Micelle assisted transfer

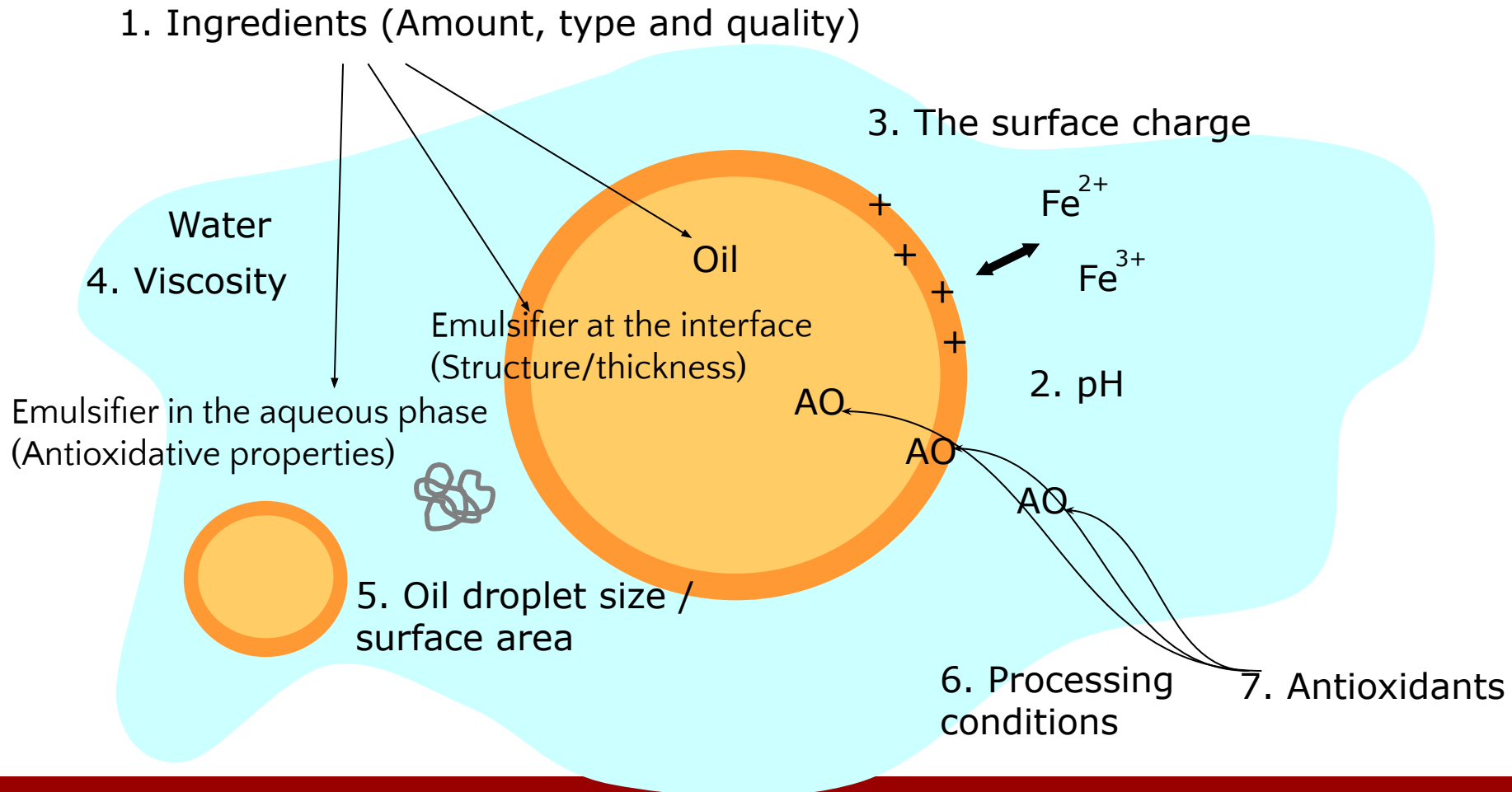


 : Micelles

*Adapted from Laguerre et al., Annual Rev. Food Sci. Technol., 2017*

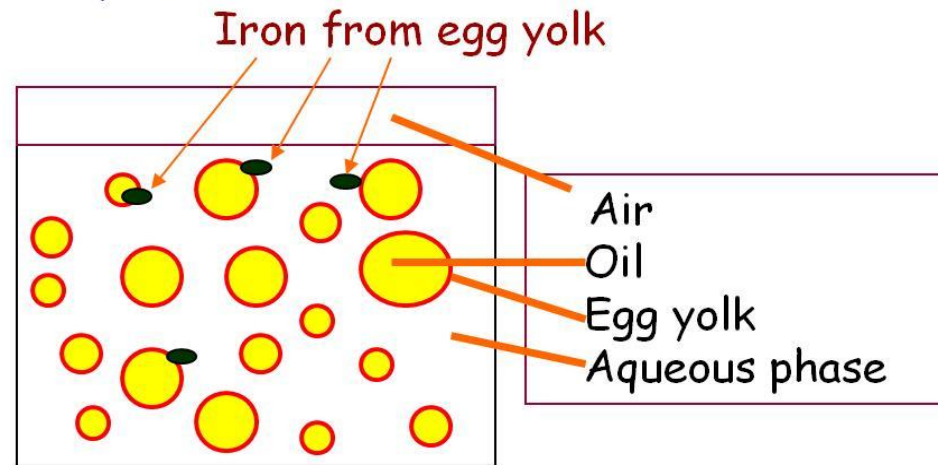
*With courtesy of P. Villeneuve, CIRAD*

# Factors that can affect lipid oxidation in emulsified foods:



# Oxidation mechanisms in mayonnaise

Mayonnaise with pH < 4



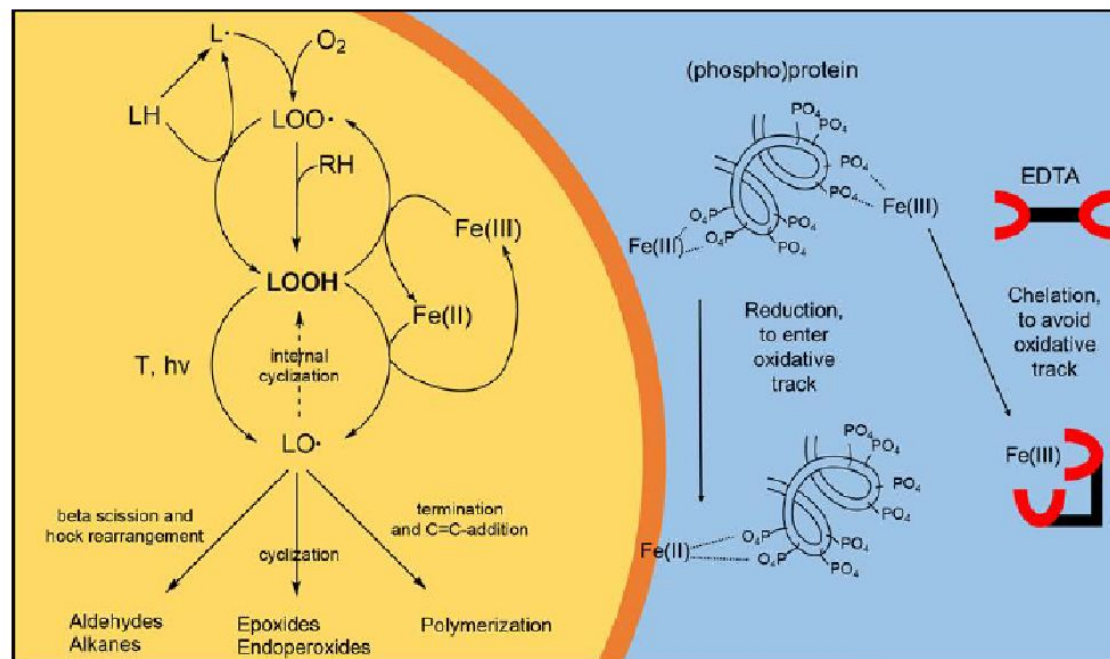
Fe in egg yolk is bound to phosvitin and lipovitilnin

- Low pH and iron from egg yolk cause oxidation in mayonnaise

EDTA is a strong antioxidant in mayonnaise because it can chelate metals and prevent oxidation from being initiated at the **interface**

*Jacobsen, PhD thesis 1999*

# Further on the oxidation mechanism of mayonnaise



**Figure 7.2.** Schematic description of the lipid oxidation pathways in oil (left), emphasizing the central role of the hydroperoxides (LOOH) and the catalytic function of iron, a transition metal. On the right, the interplay between iron and a phosphoprotein and a chelator (EDTA) is schematically depicted. Here, unchelated iron is able to enter the oxidative track, whereas chelated iron stays mainly in the water layer, away from the oxidative interface.

*Donny Merx, PhD thesis, Wageningen University, 2021*

# Peptides and protein hydrolysates as antioxidants – a new approach



# Predicted antioxidant peptides

- AnOxPePred tool
  - Method for predicting both the **free radical scavenging** and **chelating properties** of peptides based on the collected data from previous studies
    - uses **deep learning to predict**
    - trained on a **curated dataset consisting of experimentally-tested antioxidant and non-antioxidant peptides**
  - The predicted antioxidant activity is based on the peptide amino acid sequence,
    - inherent properties (e.g. size, local structure and charge)

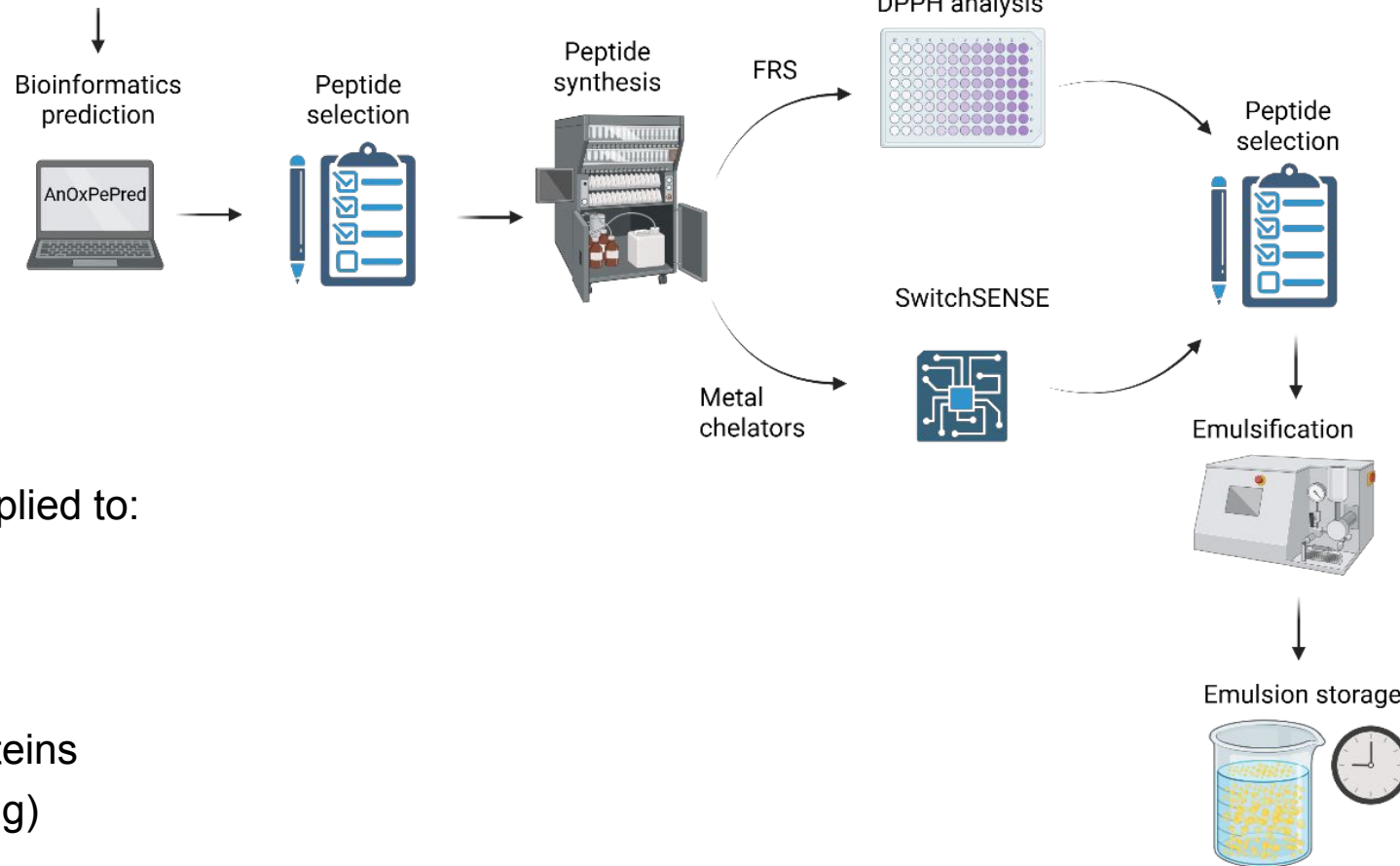
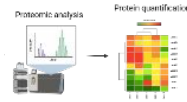
**OPEN** AnOxPePred: using deep learning for the prediction of antioxidative properties of peptides

Tobias Hegelund Olsen<sup>1</sup>, Betül Yesiltas<sup>2</sup>, Frederikke Isa Marin<sup>1</sup>, Margarita Pertseva<sup>1</sup>, Pedro J. Garcia-Moreno<sup>2</sup>, Simon Gregersen<sup>3</sup>, Michael Toft Overgaard<sup>3</sup>, Charlotte Jacobsen<sup>4</sup>, Ole Lund<sup>5</sup>, Egon Bech Hansen<sup>6</sup> & Paolo Marcatili<sup>1,2</sup>

Olsen, T.H., Yesiltas, B., Marin, F.I. *et al.* *Sci Rep* 10, 21471 (2020).  
<https://doi.org/10.1038/s41598-020-78319-w>

# Workflow Antioxidants peptides

Proteomics analysis and identification of peptides



This approach has been applied to:

- Potato proteins
- Seaweed proteins
- Single cell proteins
- Brewers spent grain proteins
- Insect proteins (upcoming)

# Peptide sources

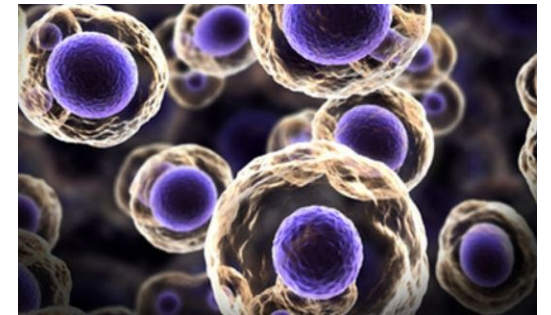
- Potato proteins
  - Obtained from potato juice, by-product in the potato starch industry
  - Protein fraction of juice at 1-2%
- Seaweed proteins from carrageenan production
  - Global annual production at  $6.5 \times 10^6$  tons
  - Protein fraction at 10-47% of the dry weight
- Single cell proteins
  - Edible unicellular microorganisms can be obtained from fermentation of methane
  - High production control and protein yield (70%)



potatoes

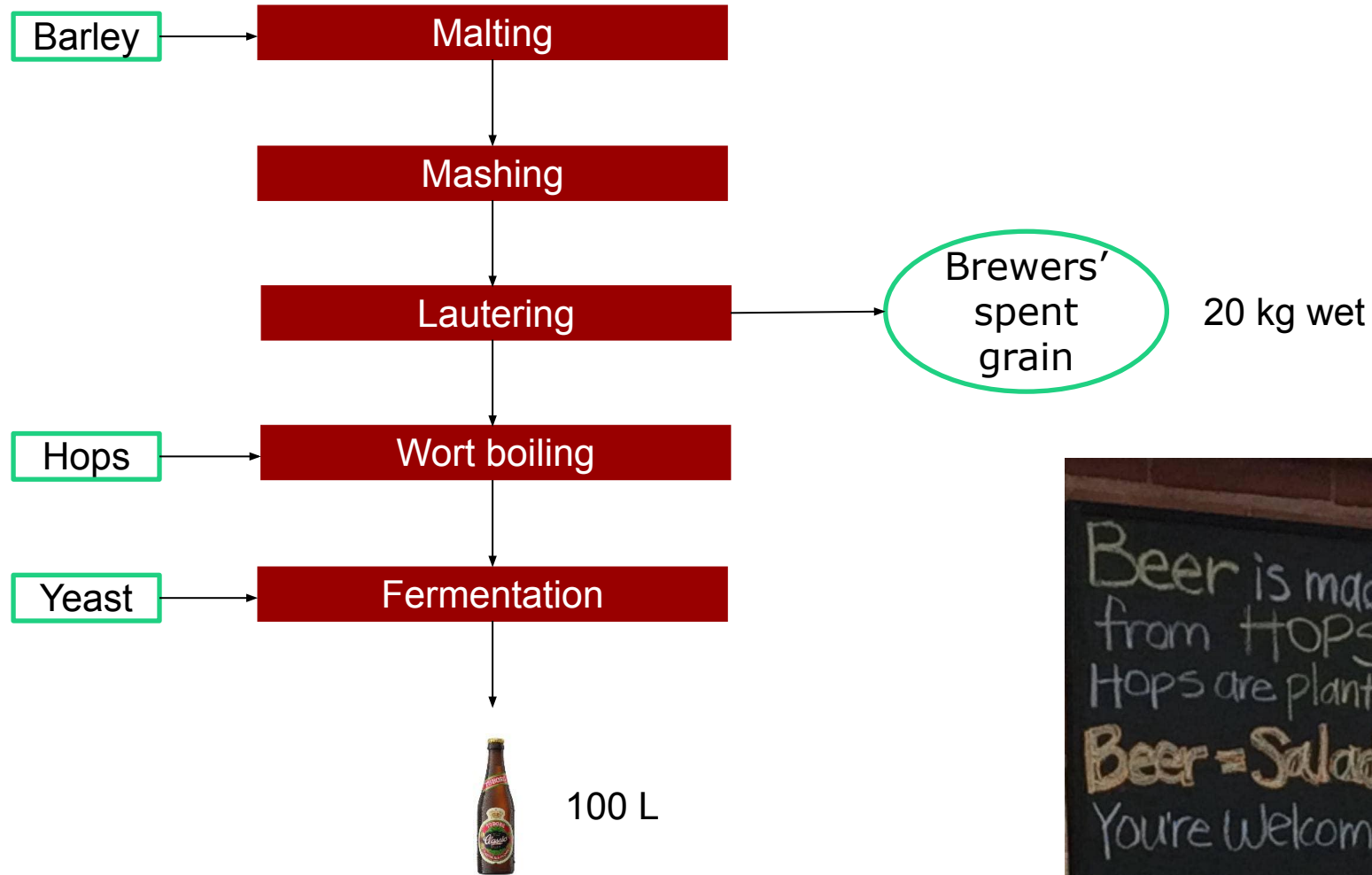


Red seaweed



single cell

# Simplified brewing process



<https://brewerywastewater.com/spent-grain/>



# Peptides evaluated in emulsions at pH7

# Selected antioxidant peptides

E1 – control

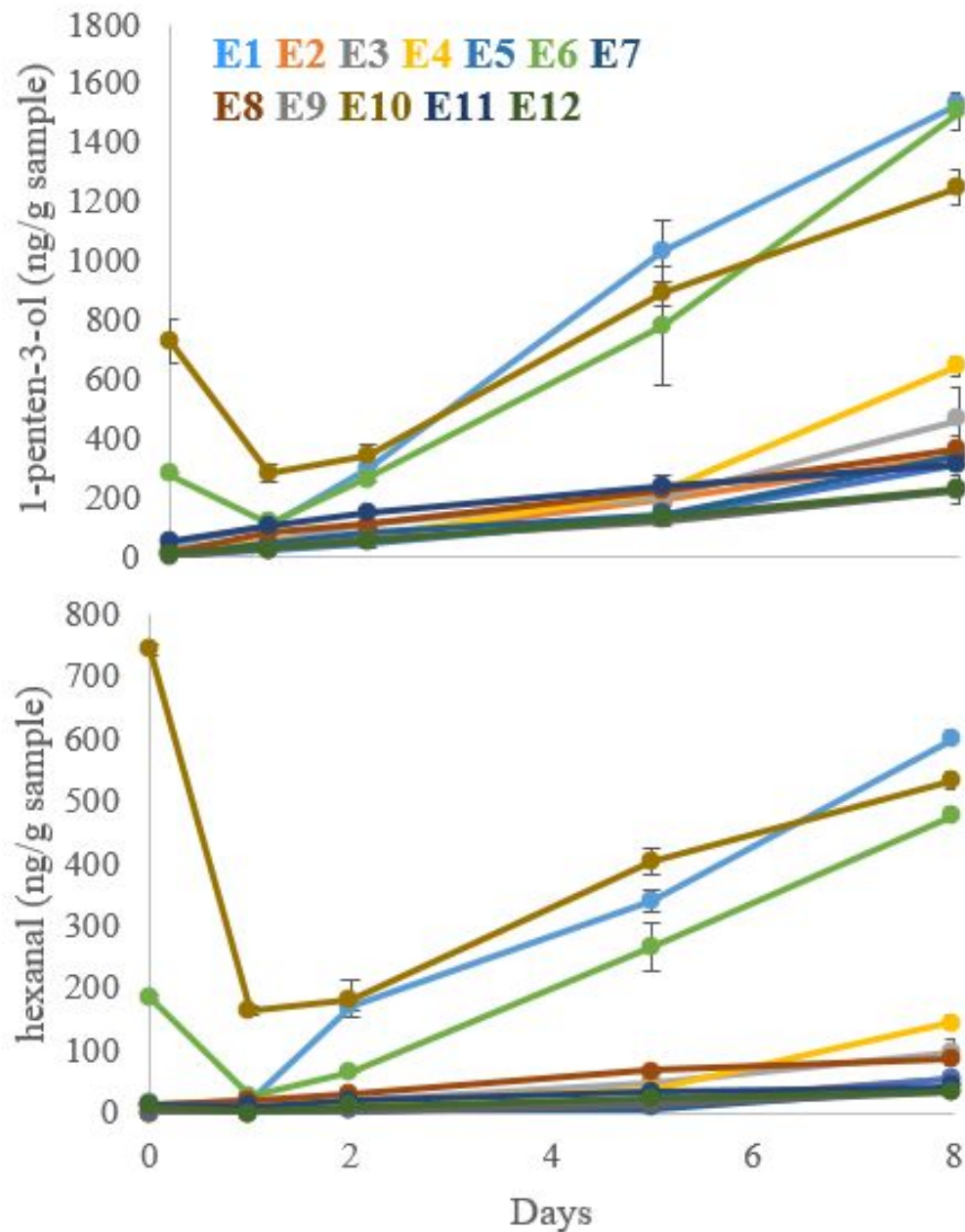
| Peptide No | Amino acid sequence    | Length | pI   | Net charge at pH7 | PRS score | PMC score | DPPH IC <sub>50</sub> (mg/ml) | Relative abundance | Tryptic match | Emulsion code |
|------------|------------------------|--------|------|-------------------|-----------|-----------|-------------------------------|--------------------|---------------|---------------|
| 111-P-SCA  | KWGPLRW                | 7      | 11.4 | 2                 | 0.46      | 0.22      | 6±2                           | 11.2% / 14.1%      | Moderate      | E2            |
| 113-P-SCA  | VPFYFEHGPHI            | 11     | 6.1  | -0.8              | 0.64      | 0.22      | 16±2                          | 23.2% / 18.6%      | Terrible      | E3            |
| 123-S-SCA  | DFPVR                  | 5      | 6.7  | 0                 | 0.38      | 0.26      | 4±1                           | 27.1% / 0.7%       | Good/Perfect  | E4            |
| 124-S-SCA  | AGDWLIGDR              | 9      | 3.7  | -1                | 0.43      | 0.20      | 3±0                           | 2.7% / 1.9%        | Perfect       | E5            |
| 125-U-SCA  | HWYD                   | 4      | 4.9  | -0.9              | 0.59      | 0.23      | 2±0                           | 7.5% / 4.1%        | Moderate      | E6            |
| 128-U-SCA  | MLWQYKPK               | 8      | 10.2 | 2                 | 0.54      | 0.20      | 7±2                           | 7.3% / 8.4%        | Good/Perfect  | E7            |
| 132-R-SCA  | NNKWVPCLEFETEHGFVYREHH | 22     | 5.8  | -1.8              | 0.50      | 0.23      | 5±2                           | 18.2%              | Moderate      | E8            |
| 133-R-SCA  | YWTMWK                 | 6      | 9.5  | 1                 | 0.53      | 0.20      | 14±1                          | 18.2%              | Perfect       | E9            |
| 135-P-CHE  | HCPSH                  | 5      | 7.2  | 0.1               | 0.51      | 0.30      |                               | 17.7% / 11.9%      | Moderate      | E10           |
| 139-P-CHE  | YKLLHCPSHLQCKN         | 14     | 8.7  | 2.1               | 0.47      | 0.26      |                               | 17.7% / 11.9%      | Good          | E11           |
| 144-P-CHE  | DDDNLVLPEDYDQD         | 14     | 0.5  | -6                | 0.45      | 0.30      |                               | 17.7% / 11.9%      | Bad/Terrible  | E12           |

**Antioxidant peptides** (0.05 wt%) were added in **5% fish oil-in-water emulsions** stabilized with 1 wt% Tween 20

-Microfluidizer □ homogenization; 50 µl Fe<sup>2+</sup> □ initiate oxidation

-Emulsions stored for 8 days in darkness





## Volatile compounds

- Similar trend was observed for the control E1 and E6 and E10
  - oxidized more than the others
  - in line with PV and toco. results
- Most stable emulsions were observed for E12, E11, and E9, followed by E2, E5 and E7

# Evaluation of the results

| Peptide No | Amino acid sequence                                                          | Length | pI   | Net charge at pH7 | PRS score | PMC score | DPPH IC <sub>50</sub> (mg/ml) | Emulsion code |
|------------|------------------------------------------------------------------------------|--------|------|-------------------|-----------|-----------|-------------------------------|---------------|
| 111-P-SCA  | KW <b>G</b> PLRW                                                             | 7      | 11.4 | 2                 | 0.46      | 0.22      | 6±2                           | E2            |
| 113-P-SCA  | VPFYFEHGPHI                                                                  | 11     | 6.1  | -0.8              | 0.64      | 0.22      | 16±2                          | E3            |
| 123-S-SCA  | DFPVR                                                                        | 5      | 6.7  | 0                 | 0.38      | 0.26      | 4±1                           | E4            |
| 124-S-SCA  | A <b>G</b> DW <b>L</b> I <b>G</b> DR                                         | 9      | 3.7  | -1                | 0.43      | 0.20      | 3±0                           | E5            |
| 125-U-SCA  | HWYD                                                                         | 4      | 4.9  | -0.9              | 0.59      | 0.23      | 2±0                           | E6            |
| 128-U-SCA  | M <b>L</b> W <b>Q</b> Y <b>K</b> P <b>K</b>                                  | 8      | 10.2 | 2                 | 0.54      | 0.20      | 7±2                           | E7            |
| 132-R-SCA  | NNKWVPCLEFETEHGFVYREHH                                                       | 22     | 5.8  | -1.8              | 0.50      | 0.23      | 5±2                           | E8            |
| 133-R-SCA  | Y <b>W</b> T <b>M</b> W <b>K</b>                                             | 6      | 9.5  | 1                 | 0.53      | 0.20      | 14±1                          | E9            |
| 135-P-CHE  | HCPSH                                                                        | 5      | 7.2  | 0.1               | 0.51      | 0.30      | -                             | E10           |
| 139-P-CHE  | Y <b>K</b> <b>L</b> L <b>H</b> C <b>P</b> S <b>H</b> L <b>Q</b> C <b>K</b> N | 14     | 8.7  | 2.1               | 0.47      | 0.26      | -                             | E11           |
| 144-P-CHE  | D <b>D</b> D <b>N</b> L <b>V</b> L <b>P</b> E <b>V</b> D <b>Q</b> D          | 14     | 0.5  | -6                | 0.45      | 0.30      | -                             | E12           |

• Better performing peptides are rich in antioxidant and hydrophobic AAs

- **antioxidant:** W (tryptophan), Y (tyrosine), D (aspartic acid), M (methionine), R (arginine), H (histidine) and K (lysine)
- **hydrophobic:** A (alanine), F, (phenylalanine, G (glycine), V (valine), P (proline) and L (leucine)

# Peptides selected for evaluation in emulsions and mayonnaise based on AnOxPePred and *in vitro* radical scavenging assay

## Study A

**5% fish o/w emulsion  
at pH4**

- Physical and oxidative stability
- 11 peptides selected from radical scavenger assay

## Study B

**Mayonnaise**

- Oxidative stability of peptides
- Best performing 4 peptides in Study A

## Study C

**Mayonnaise**

- Concentration effect on oxidative stability
- Best performing peptide in Study B

# Study A - 5% fish o/w emulsion at pH4

| Emulsion code | Peptide No | Amino acid sequence        | pI    | PRS score* | PC score* |
|---------------|------------|----------------------------|-------|------------|-----------|
| E2            | 111-P-SC A | KWGPLRW                    | 11.39 | 0.46       | 0.22      |
| E4            | 123-S-SC A | DFPVR                      | 6.68  | 0.38       | 0.26      |
| E6            | 125-U-SC A | HWYD                       | 4.87  | 0.59       | 0.23      |
| E8            | 132-R-SC A | NNKWVPCLEFET<br>EHGFVYREHH | 5.79  | 0.50       | 0.23      |

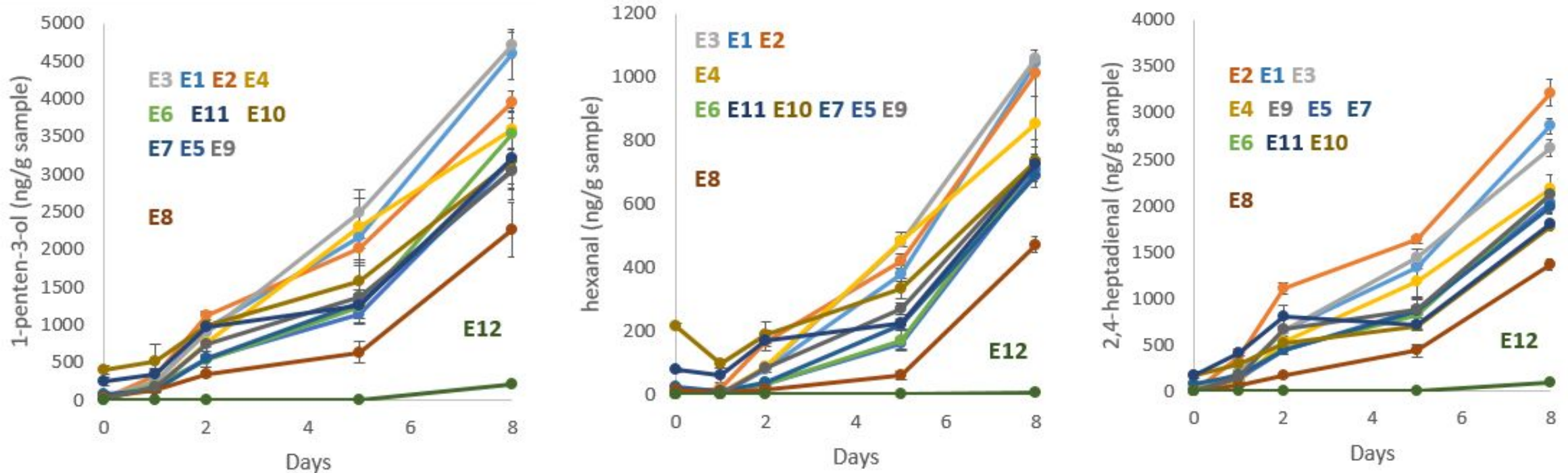
Antioxidant peptides (0.05 wt%) were added in 5% fish oil-in-water emulsions stabilized with 1 wt% Tween 20 at pH 4

- Microfluidizer □ homogenization;  
50 µl Fe<sup>2+</sup> □ initiate oxidation

- pH ~4
- D[3,2] □ 131 - 136 nm
- D[4,3] □ 227 - 244 nm
- ζ-potential □ (-) 4.7 - (-) 2.0 mV

# Study A - 5% fish o/w emulsion at pH4

## Oxidative stability results – volatile compounds



- Three times higher oxidation levels compared to pH7 emulsions
- **E12** had the lowest development of volatiles during storage, followed by **E8**. **At pH 7 E8 was poor.**
- Control and **E2** was the worst and results were mostly in line with the PV and tocopherol results. **At pH7 E2 was good**

*Varona et al. Food Chem, 2023*

# Study B – Mayonnaise with antioxidant peptides

## Experimental set up and storage

- Fish oil enriched mayonnaise (20% of the total oil replaced with fish oil)
- 4 peptides (75 ppm) tested, one from each protein source
  - Seaweed - 124-S-SCA (E4)
  - Single cell - 128-U-SCA (E7)
  - Spinach - 132-R-SCA (E8)
  - Potato - 144-P-CHE (E12)
  - EDTA (commonly used synthetic antioxidant)
- Control mayonnaise without any antioxidant added
- 4 weeks of storage at RT, in darkness

| PEPTIDES  | Sequence                | pI    |
|-----------|-------------------------|-------|
| 124-S-SCA | AGDWLIGDR               | 3.71  |
| 128-U-SCA | MLWQYKPK                | 10.21 |
| 132-R-SCA | NNKWVPCLEFETEHGTFVYREHH | 5.79  |
| 144-P-CHE | DDDNLVLPEVYDQD          | 0.52  |

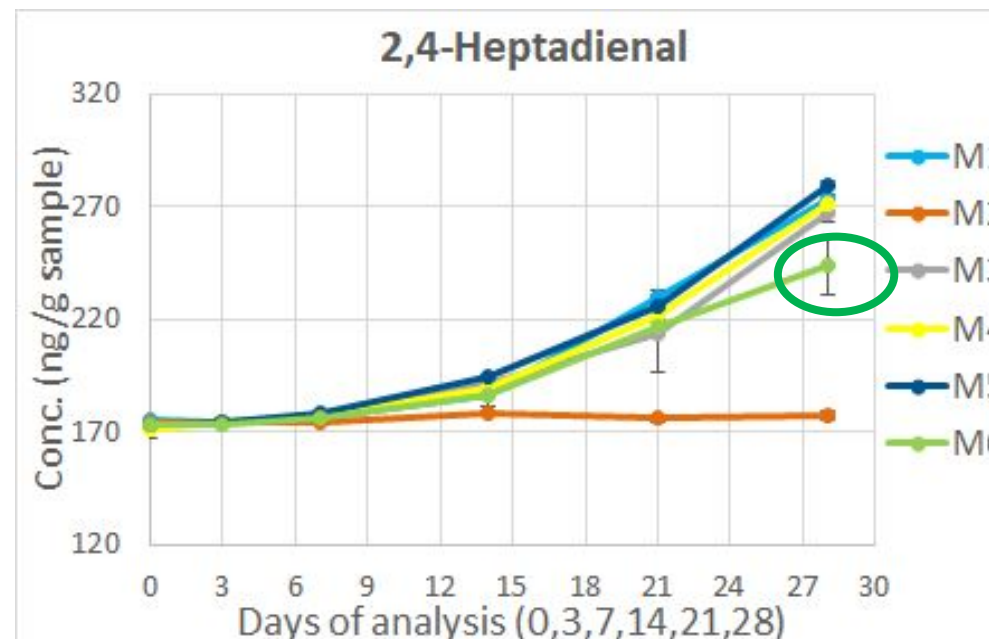
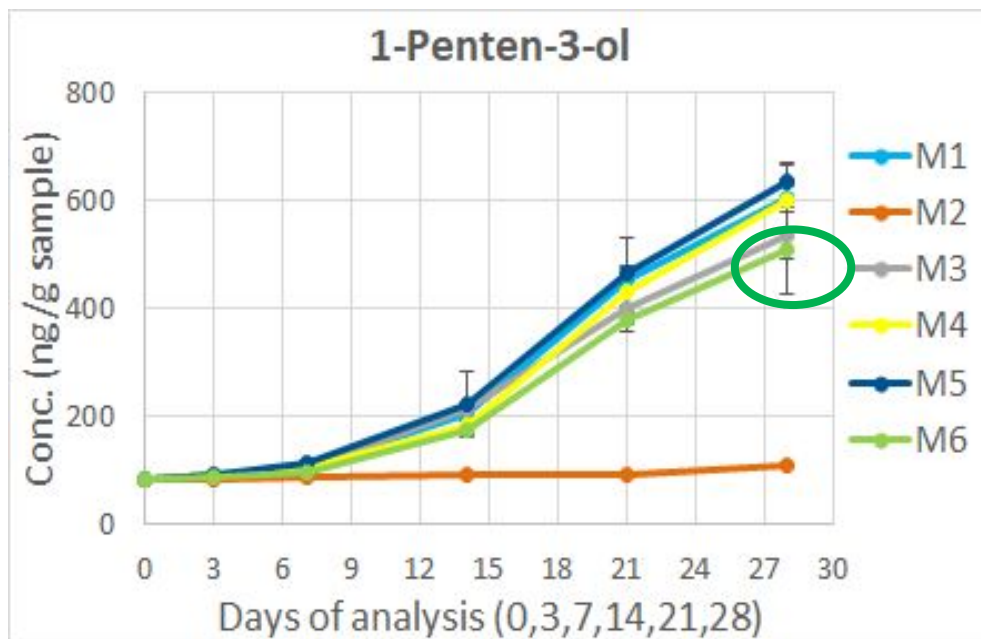
| Emulsion code | Antioxidant |
|---------------|-------------|
| M1            | -           |
| M2            | EDTA        |
| M3            | 124-S-SCA   |
| M4            | 128-U-SCA   |
| M5            | 132-R-SCA   |
| M6            | 144-P-CHE   |





# Study B – Mayonnaise with antioxidant peptides

## Oxidative stability – volatile compounds



□ **EDTA** had a very low volatile compound formation

□ **Potato** peptide had the lowest levels among other peptides (E12)

□ **Potato** peptide (144-CHE selected for concentration studies)

**Note that it was predicted to be a metal chelator!**

| Emulsion code | Antioxidant |
|---------------|-------------|
| <b>M1</b>     | -           |
| <b>M2</b>     | EDTA        |
| <b>M3</b>     | 124-S-SCA   |
| <b>M4</b>     | 128-U-SCA   |
| <b>M5</b>     | 132-R-SCA   |
| <b>M6</b>     | 144-P-CHE   |

*Varona et al. Food Chem, 2023*

# Study C – Concentration effect of peptide in mayonnaise

## Experimental set up and storage

### Best performing antioxidant potato peptide

| Name      | Sequence       | Length | pI   |
|-----------|----------------|--------|------|
| 144-P-CHE | DDDNLVLPEDVDQD | 14     | 0.52 |



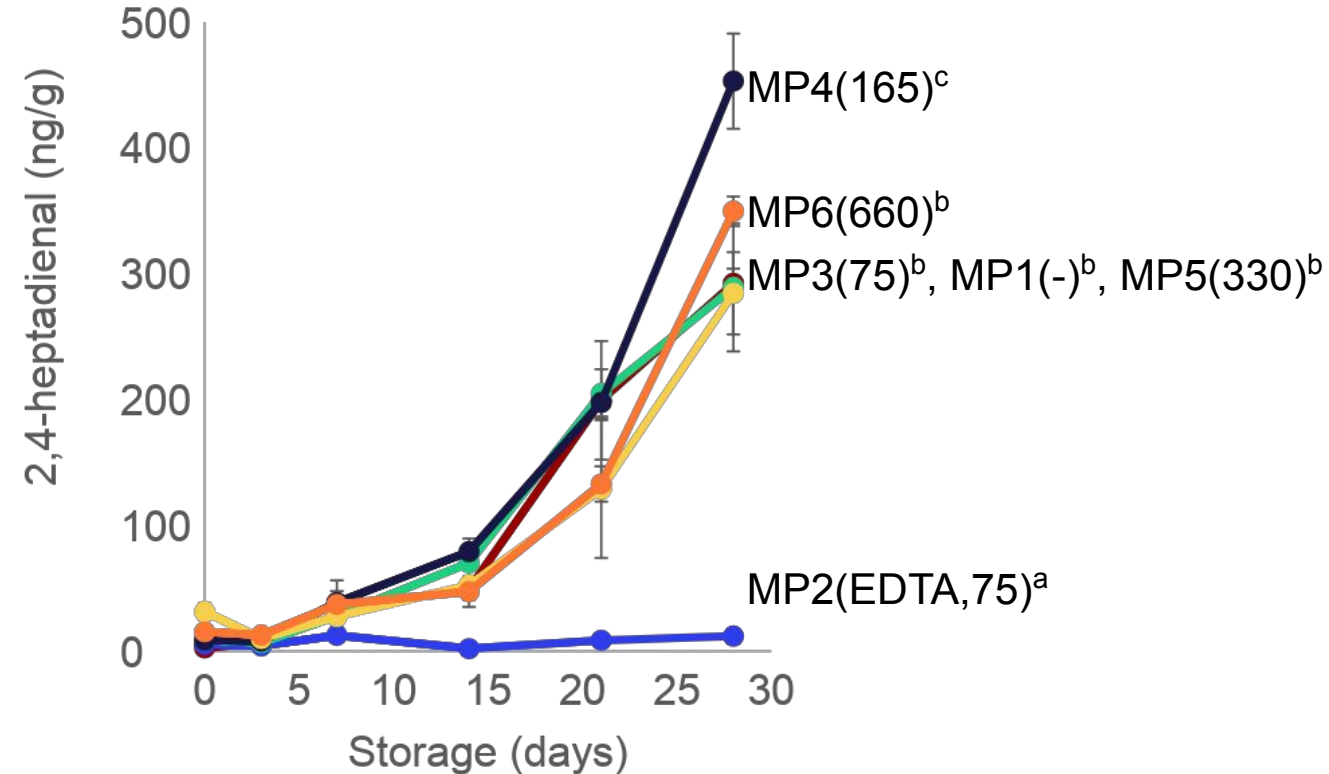
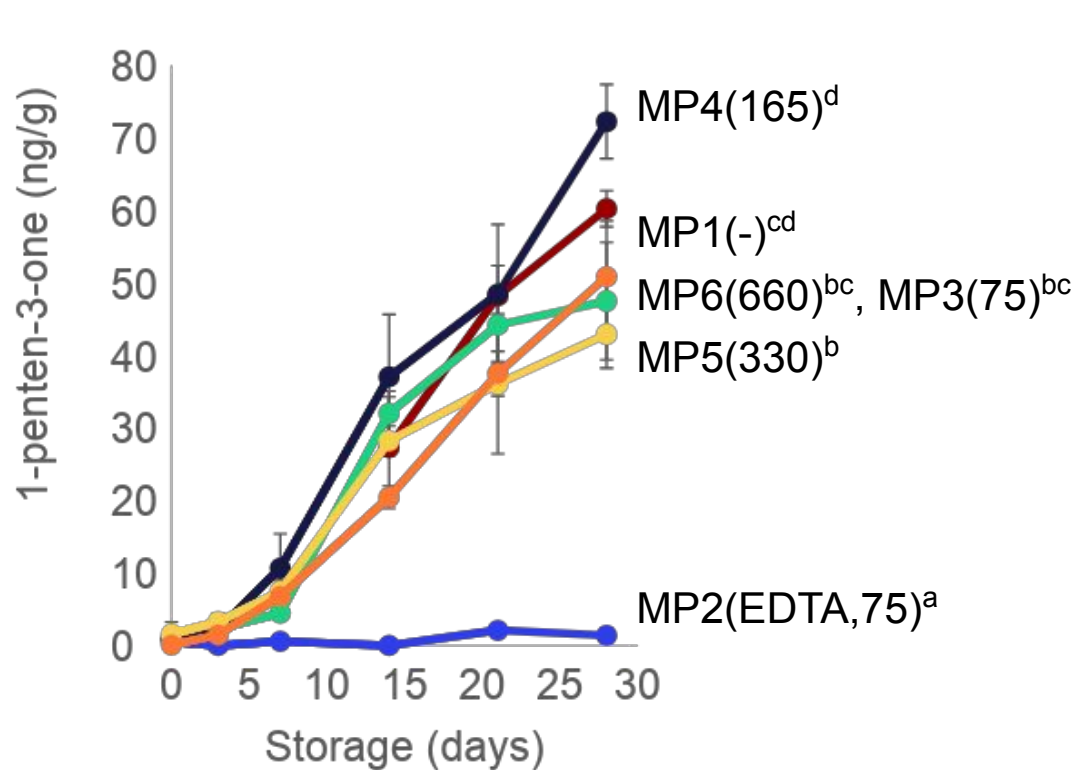
### 28-day fish oil enriched mayonnaise storage experiment at RT

| Mayonnaise code | Antioxidant used | Antioxidant, ppm |
|-----------------|------------------|------------------|
| MP1 (-)         | -                | -                |
| MP2 (EDTA,37.5) | EDTA             | 75               |
| MP3 (37.5)      | 144-P-CHE        | 75               |
| MP4 (82.6)      | 144-P-CHE        | 165              |
| MP5 (165.3)     | 144-P-CHE        | <b>330</b>       |
| MP6 (330.6)     | 144-P-CHE        | 660              |

- Control mayo without any antioxidants, **MP1**
- Mayo with EDTA, **MP2**
- Mayo with potato peptide, **MP3-6**
  - Peptide concentrations starting with the same mass concentration as for EDTA and increased up to the **equimolar concentration of EDTA** and twice as much

# Study C – Concentration effect of peptide in mayonnaise

## Oxidative stability – volatile compounds



- **EDTA** provided much better results compared to potato peptides in all concentrations
- Lowest oxidation values for potato peptide was added in the same molar concentration as EDTA

**Peptides selected for evaluation in mayonnaise based on AnOxPePred scores for metal chelating activity and other parameters related to potential metal chelating activity**

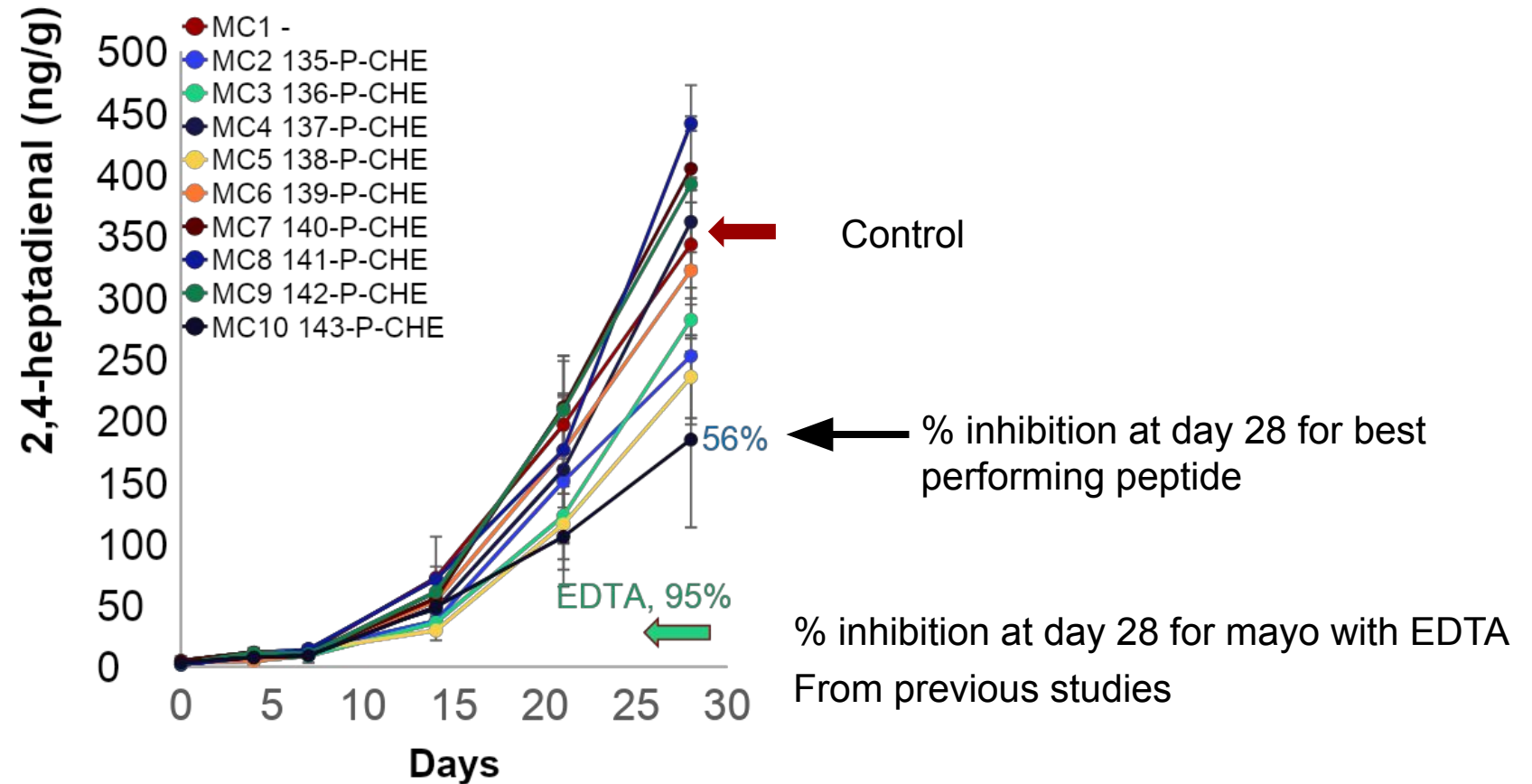
# Selected peptides



| Protein source | Sequence                     | Peptide   | Length | pI   | PFRS | PMCS | RA-A (%) | RA-B (%) | TM |
|----------------|------------------------------|-----------|--------|------|------|------|----------|----------|----|
| potato         | HCPSH                        | 135-P-CHE | 5      | 7.23 | 0.51 | 0.30 | 17.70%   | 11.90%   | 4  |
|                | MDDAS                        | 136-P-CHE | 5      | 0.64 | 0.31 | 0.23 | 13.23%   | 33.53%   | 1  |
|                | CEGES                        | 137-P-CHE | 5      | 0.86 | 0.36 | 0.23 | 1.10%    | 0.12%    | 5  |
|                | ALQWMLVI                     | 138-P-CHE | 8      | 3.72 | 0.44 | 0.20 | 16.34%   | 12.01%   | 1  |
|                | YKLLHCPSHLQCKN               | 139-P-CHE | 14     | 4.00 | 0.47 | 0.26 | 17.70%   | 11.90%   | 6  |
|                | GMKNVYKLLHCPSHLQCKNIGSN      | 140-P-CHE | 23     | 9.65 | 0.53 | 0.25 | 4.60%    | 2.00%    | 1  |
|                | LGYNLLYCPFSSDDQFCLKVGVVHQN   | 141-P-CHE | 26     | 5.03 | 0.29 | 0.15 | 6.76%    | 3.37%    | 4  |
|                | ENPIVLPSTCHDDDNLVLPVYDQDGHPL | 142-P-CHE | 29     | 3.42 | 0.35 | 0.26 | 7.81%    | 2.19%    | 6  |
|                | DFNLVDGDVAADVPSLLSISV        | 143-P-CHE | 21     | 0.42 | 0.30 | 0.23 | 2.70%    | 4.80%    | 1  |
|                | DDDNLVLPVYDQD                | 144-P-CHE | 14     | 0.52 | 0.45 | 0.30 | 17.70%   | 11.90%   | 2  |
| seaweed        | DLSDLSQP                     | 145-S-CHE | 8      | 0.73 | 0.32 | 0.29 | 2.76%    |          | 4  |
|                | QLVQLLPFHV                   | 146-S-CHE | 11     | 7.52 | 0.45 | 0.24 | 5.59%    | 0.06%    | 4  |
|                | IDLPLKQSDVDPVVAKCCEV         | 147-S-CHE | 21     | 3.61 | 0.37 | 0.14 | 2.76%    |          | 1  |
|                | QSDVDPVVAK                   | 148-S-CHE | 10     | 3.71 | 0.31 | 0.23 | 2.76%    |          | 9  |
|                | EAEMR                        | 149-S-CHE | 5      | 4.15 | 0.37 | 0.23 | 2.76%    |          | 7  |
| single cell    | PLHVPV                       | 150-U-CHE | 6      | 8.26 | 0.50 | 0.27 | 8.89%    | 32.36%   | 1  |
|                | PSLGQQAQAPA                  | 151-U-CHE | 11     | 4.07 | 0.39 | 0.24 | 1.21%    | 0.03%    | 3  |
|                | MVDALQFHEEHGEVCPAGWNK        | 152-U-CHE | 21     | 4.51 | 0.59 | 0.22 | 2.68%    | 1.41%    | 9  |
|                | CVQSHR                       | 153-U-CHE | 6      | 9.21 | 0.39 | 0.24 | 1.26%    | 0.96%    | 9  |
|                | MEWEVPEK                     | 154-U-CHE | 8      | 3.85 | 0.47 | 0.23 | 7.26%    | 8.35%    | 9  |

# Oxidative stability – Chelator potato peptides

Volatile compounds (DHS/GC-MS)



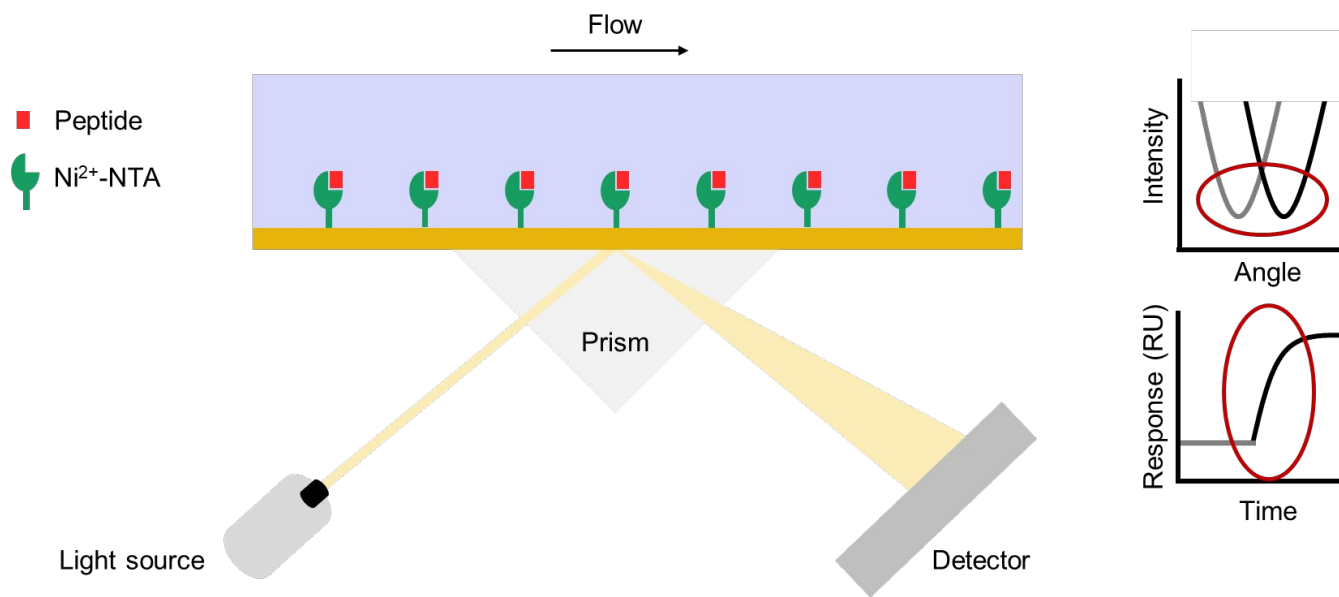


# Screening of metal chelating peptides

## Ferrozine assay does not work, so what to do?

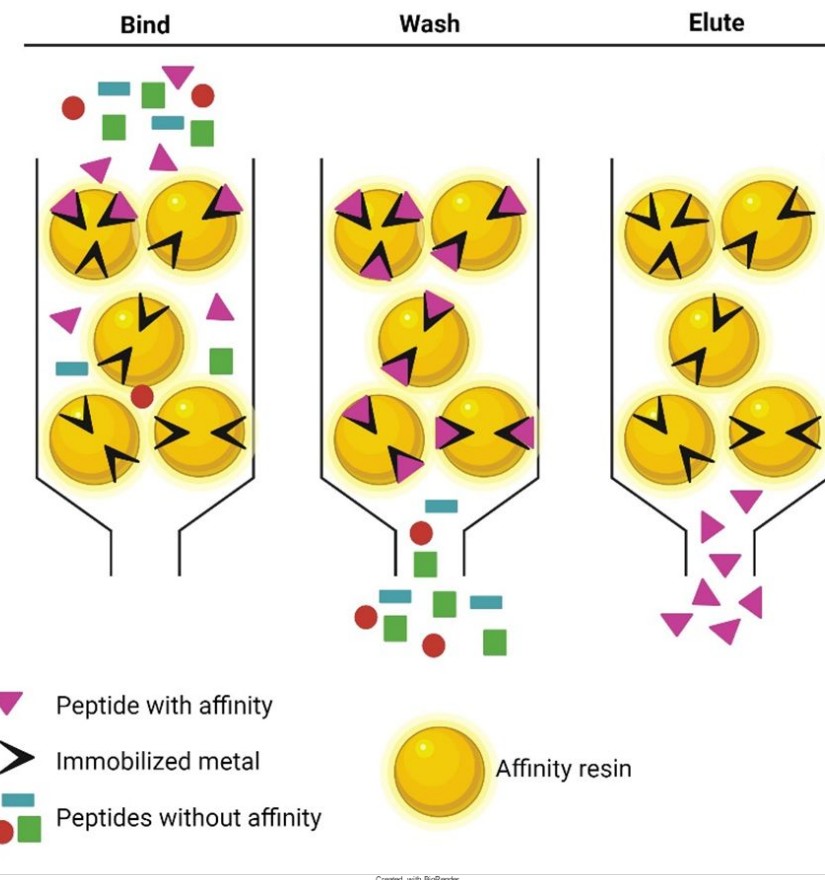
# Can we screen for metal chelating peptides?

## Examples of potential techniques

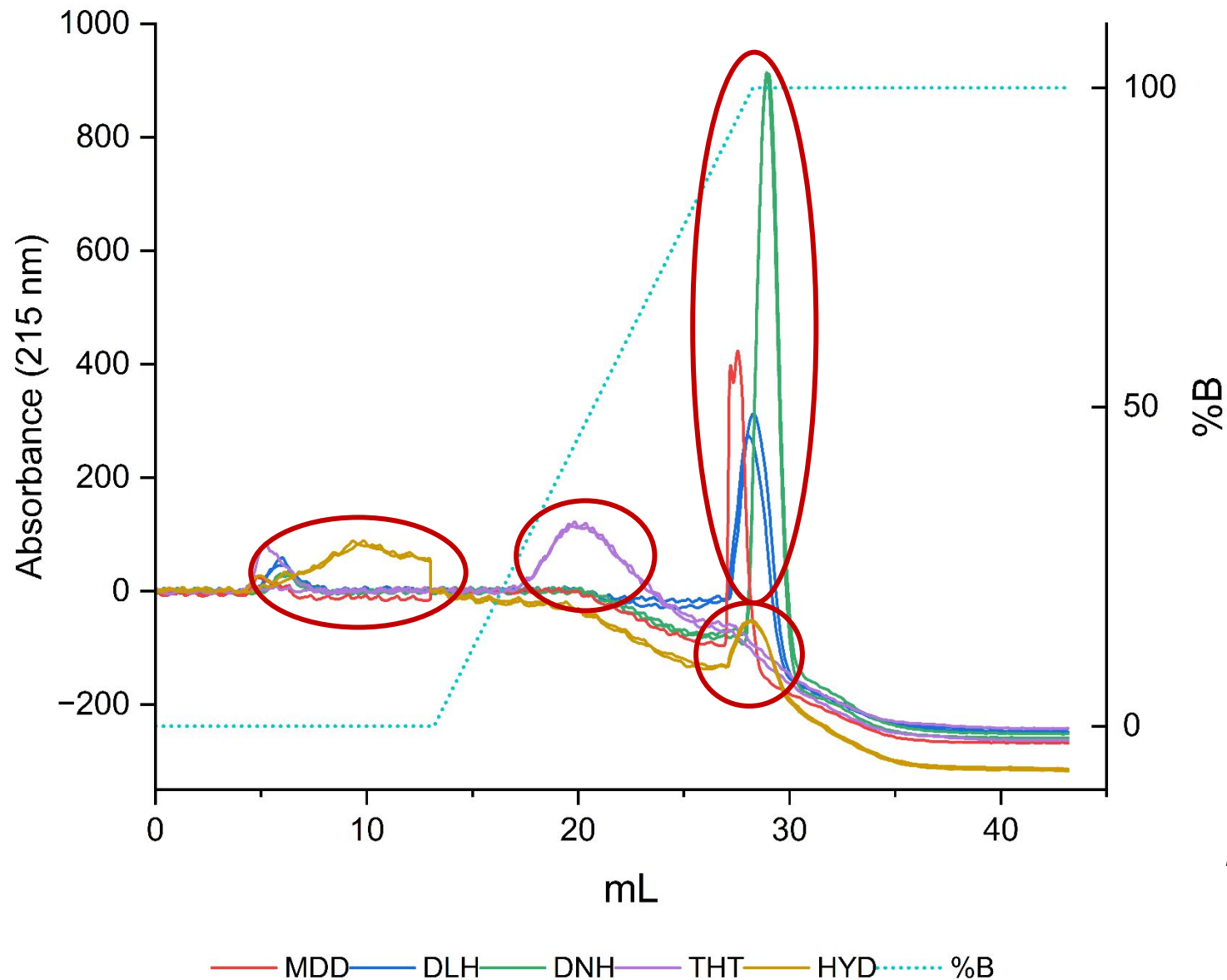


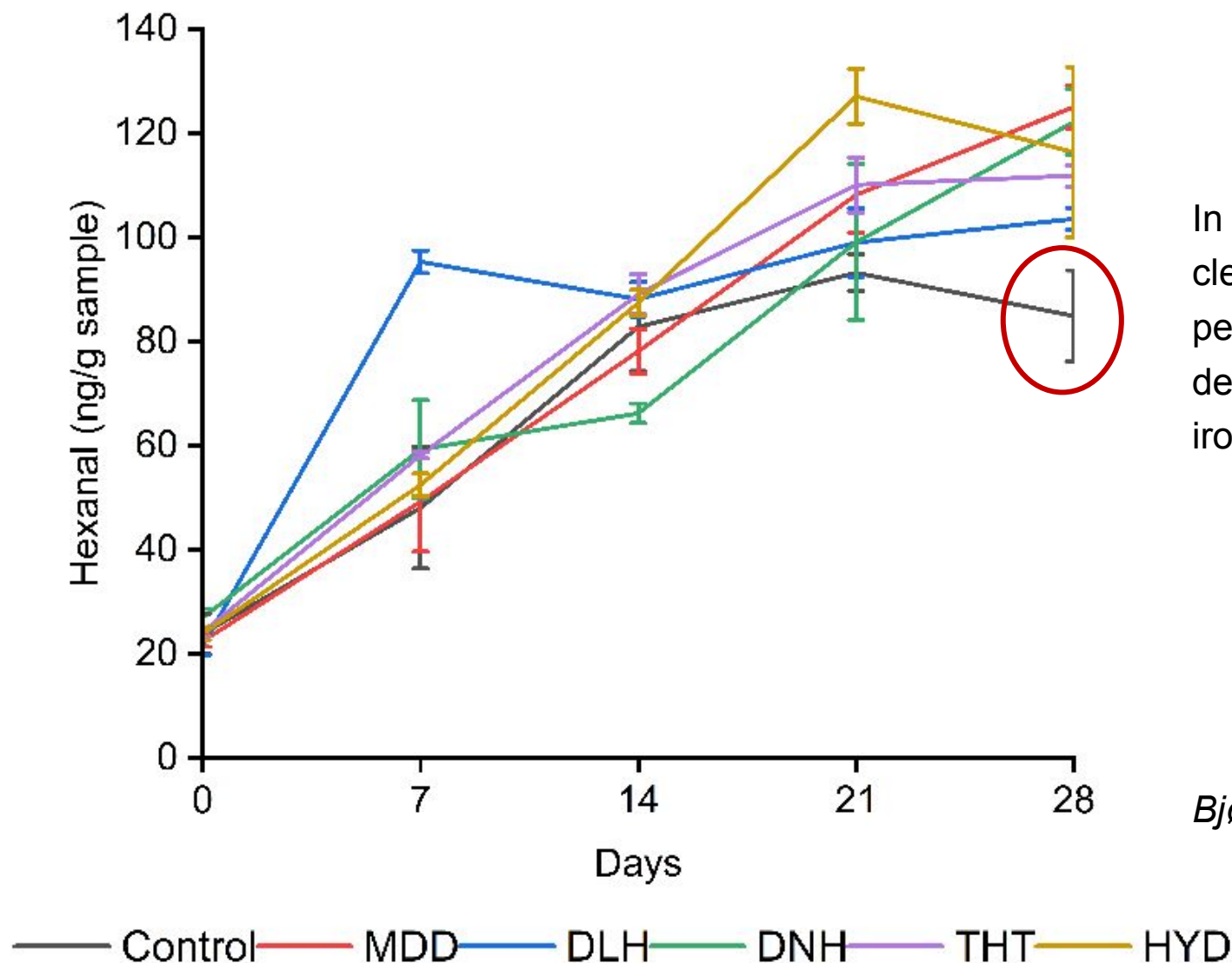
Surface plasmon resonance (SPR)

SPR works at pH7, but work by M Bjørle showed that more Development is necessary before it can be used at pH 4



Immobilized metal affinity chromatography (IMAC)

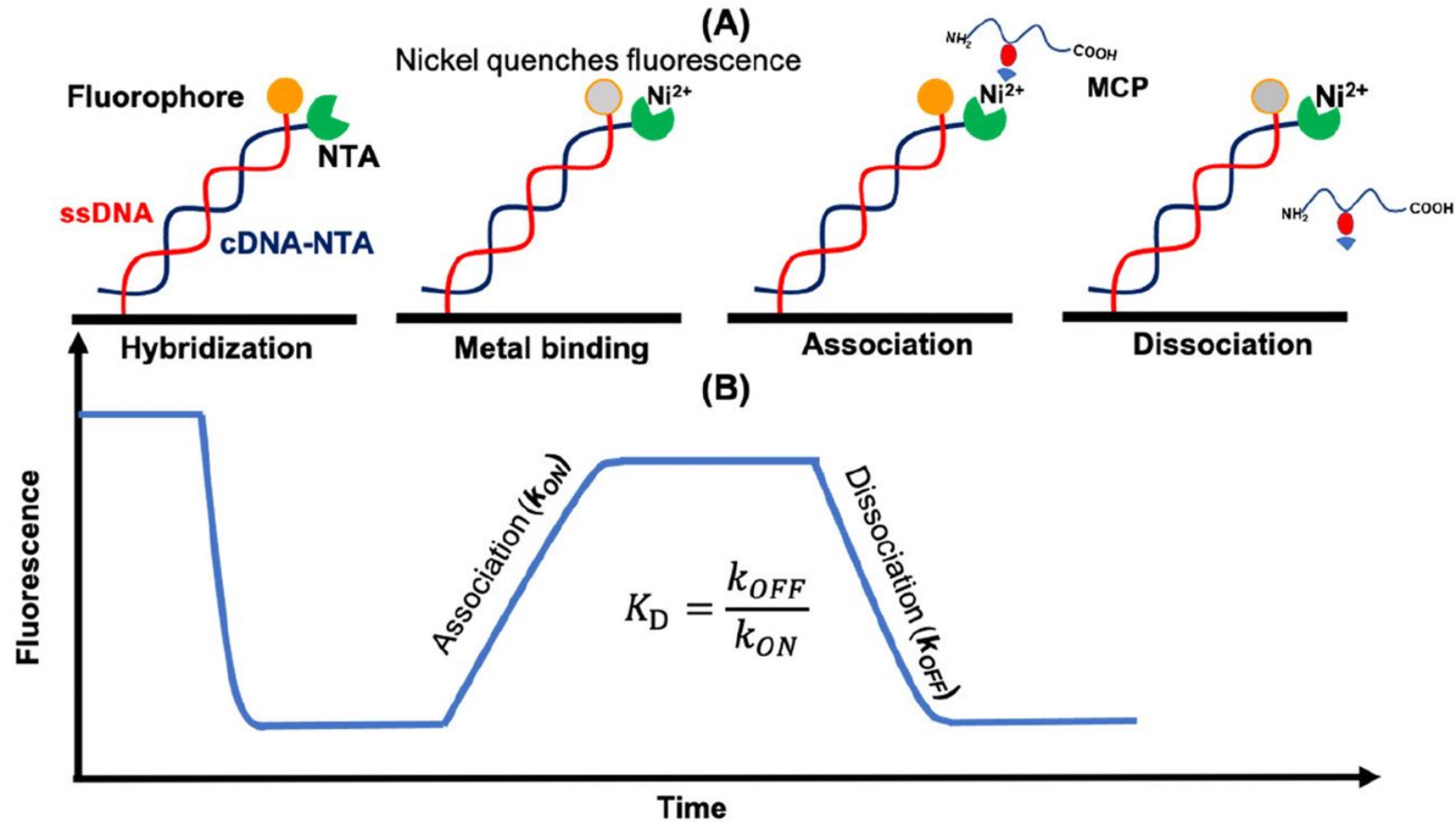
$\text{Fe}^{3+}$ *Bjørllie, M. PhD thesis 2024*



In mayonnaise there was no clear antioxidant effect of peptides and hydrolysates despite differences in iron binding capacity

*Bjørllie et al. Food Chem, 2025*

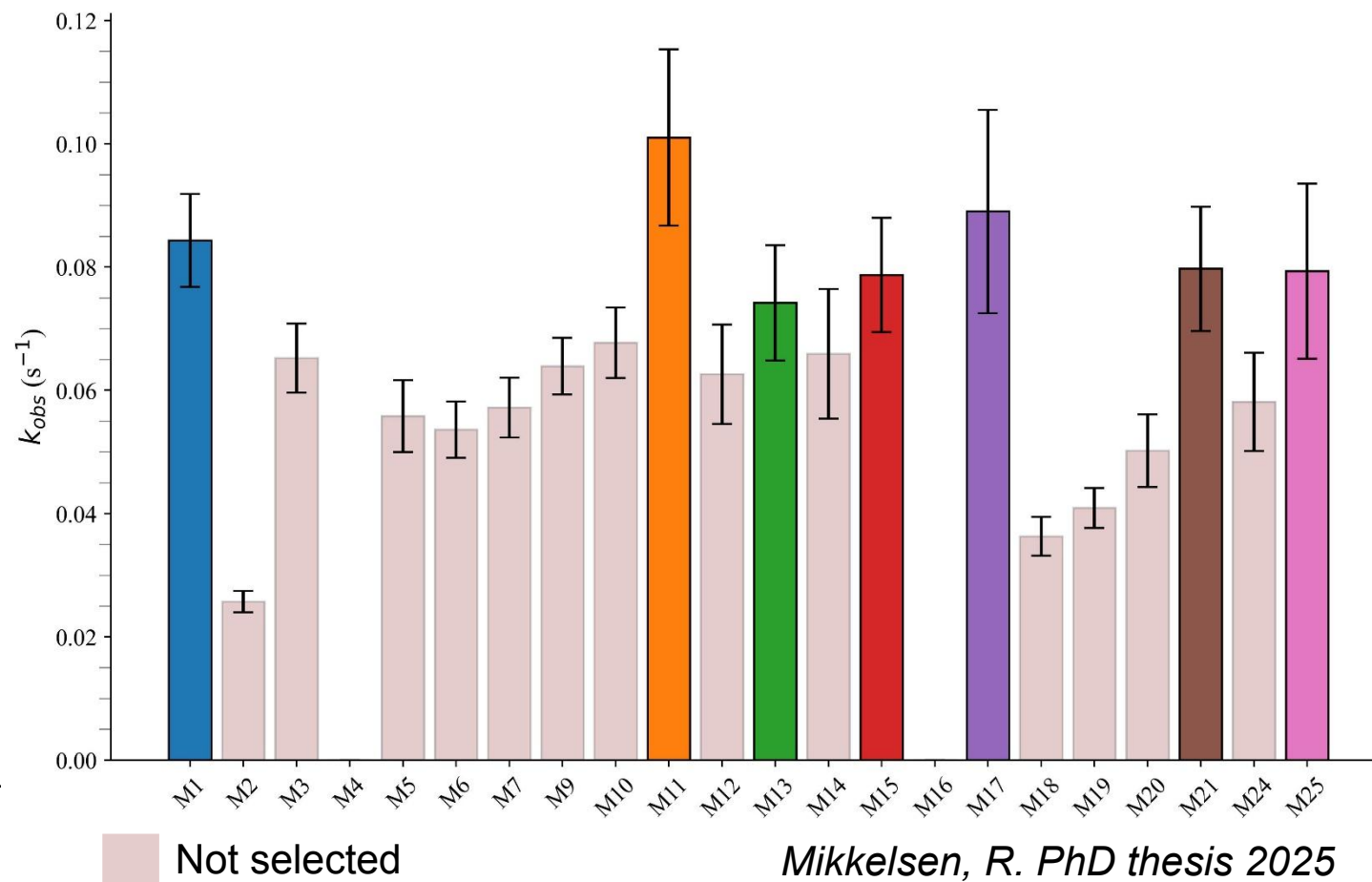
# SwitchSENSE®



# Screening of metal chelating peptides from brewers spent grain

| Metal chelating peptides |                               | AnOxPePred score |
|--------------------------|-------------------------------|------------------|
| M1                       | PQQPQPFQPPFSPQQPFPQQPPFWQQP   | 0.422            |
| M2                       | QPQFPQPPFSPQQPFPQQPPFWQQ      | 0.394            |
| M3                       | TQDQTPLLQEQQDQM               | 0.377            |
| M4                       | QVGQCSFQQPQQGQQQQ             | 0.361            |
| M5                       | TFTSSIHFLAKTHQM               | 0.343            |
| M6                       | SLQLVQGVSQPQQSQQQQ            | 0.342            |
| M7                       | PHP                           | 0.339            |
| M8                       | ESFGFSSTLRAATSGQAFPQCVFDHWD   | 0.367            |
| M9                       | FGGGLTTEQPQGGKQPFHCQQT        | 0.362            |
| M10                      | RDLHNQAQ                      | 0.361            |
| M11                      | GQQGYPTTSPQSSGQGOQLGQGQQPGQGQ | 0.358            |
| M12                      | PGSTFPQQPGQGOQPGQRQPW         | 0.354            |
| M13                      | PSTTSPQSSGQGOQLGQGQQ          | 0.353            |
| M14                      | PYPIATSPHQPGQWQQPGQGQ         | 0.353            |
| M15                      | LHPN                          | 0.350            |
| M16                      | PFLLLHRLRPNQLRPLPTT           | 0.349            |
| M17                      | DTPSSEFNHRFQELGL              | 0.349            |
| M18                      | PQQPQPFQRPFPSPQQPFPQQPPFWQQP  | 0.429            |
| M19                      | TPQQPPLPQGQQDQM               | 0.396            |
| M20                      | FPQRPFPSPQPFQ                 | 0.373            |
| M21                      | PPP                           | 0.340            |
| M22                      | EELVQQVLSAGWREGLNVACENALPRYDP | 0.110            |
| M23                      | VKLFNAVVEDLVVKENRVAGVITNWALAS | 0.080            |
| M24                      | TRRYMT                        | 0.168            |
| M25                      | MTD                           | 0.237            |

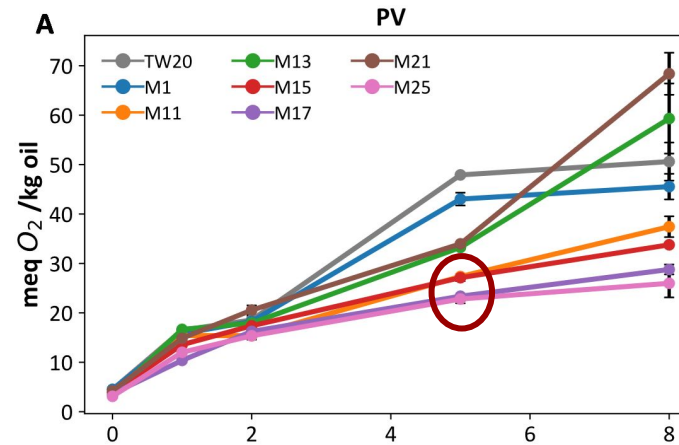
Ni association constant determined by switchSENSE®



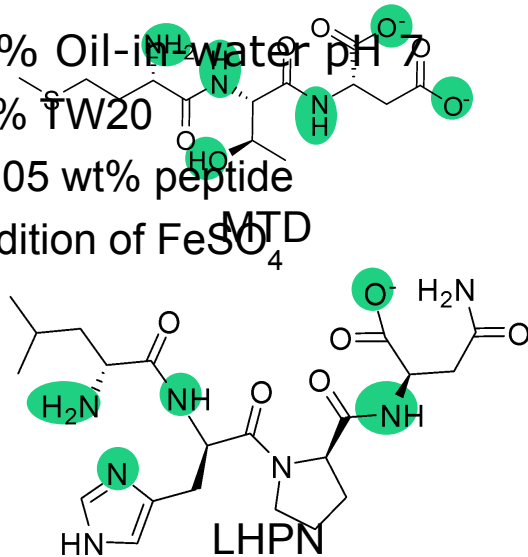
Mikkelsen, R. PhD thesis 2025

# Emulsion storage experiment

| Peptide code | Peptide Sequence              |
|--------------|-------------------------------|
| TW20         | -                             |
| M1           | PQQPQFPQQPFPSQQPFQPPFWQQQP    |
| M11          | GQQGYPSITSPQQSGQGQQLGQGQPGQGQ |
| M13          | PSTTSPQQSGQGQQLGQGQ           |
| M15          | LHPN                          |
| M17          | DTPSSEFNHRFQELGL              |
| M21          | PPP                           |
| M25          | MTD                           |



- 5% Oil-in-water pH 7
- 1% TW20
- 0.05 wt% peptide
- Addition of FeSO<sub>4</sub>



Mikkelsen, R. PhD thesis 2025



## Conclusions

- Lipid oxidation mechanisms in emulsions are complex and affected by many different factors
- Interface composition and quality of emulsifier very important for oxidative stability
- Mass transfer phenomenon not fully understood but reverse micelles from LOOH could play a role
- EDTA is still the strongest antioxidant in mayonnaise

## Conclusions

- Peptides are promising natural alternatives to EDTA especially in emulsions at neutral pH
- New screening techniques for selecting metal chelating compounds are needed
- SPR and IMAC are promising techniques but need improvement
- Switchsense has also shown promising results

# Acknowledgements

- Former and current members of the Bioactives – Analysis and Application Research Group
- Industrial collaborators
- Collaborators in the PROVIDE and GRAINPEP projects





# **WORLD CONGRESS ON OILS & FATS AND ISF LECTURESHIP SERIES**

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